

# **ADVANCES IN THE USE OF TROPHOBLASTIC CELLS FOR PRENATAL NON-INVASIVE DIAGNOSTICS OF GENETIC DISORDERS**

**15th INTERNATIONAL CONFERENCE  
ON PREIMPLANTATION GENETIC DIAGNOSIS  
BOLOGNE 8-11 May 2016**

**Patrizia Paterlini Bréchet, MD, Ph.D.  
Professor of Cellular and Molecular Biology  
University Paris Descartes– Paris**



## **Disclosure**

**Inventor/coinventor of ISET patents**

**Founder and scientific advisor of Rarecells**

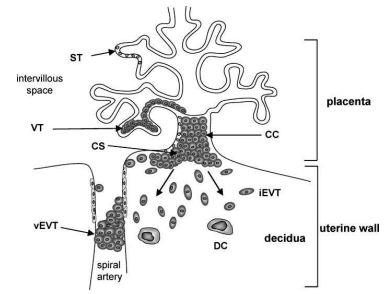
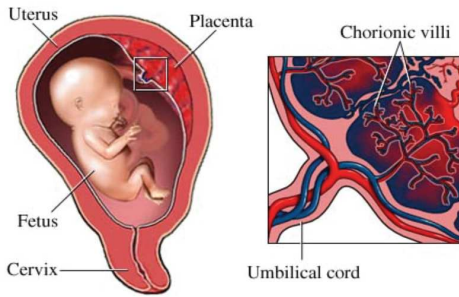
**Academic tasks:**

**Teaching (University Paris Descartes)**

**Developing and implementing new tests (Hôpital Necker)**

**Research activity (INSERM Unit)**





## Circulating Fetal Trophoblastic Cells (CFTC)

**CFTC do not have to cross the placenta**

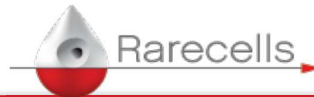
Endovascular trophoblasts invade the lumen of spiral arteries coming in contact with maternal blood

Invasion follows two waves: middle of 1T and end of 1T

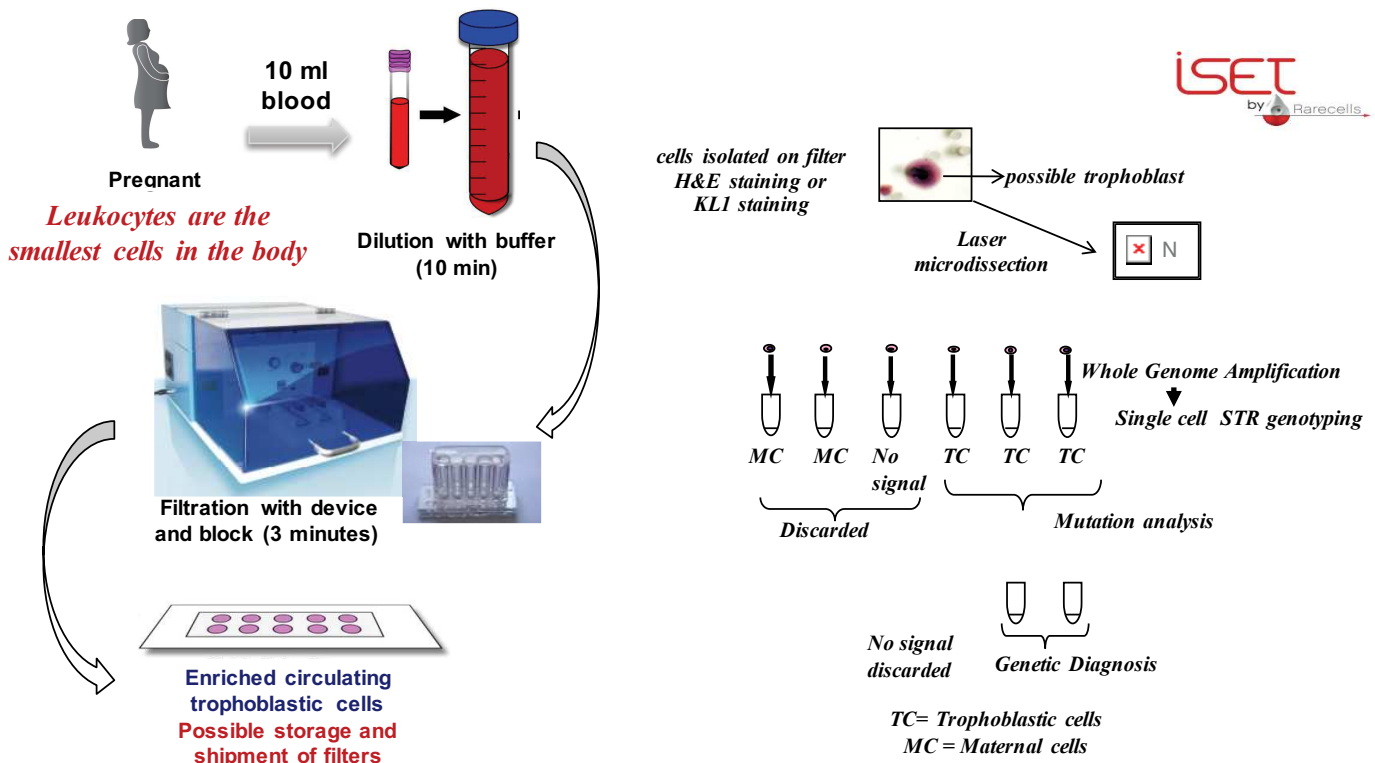
• Isolation by density gradient, MACS, from 20 to 50 ml of blood, followed by Y PCR or FISH: very inconsistent results (Oudejans C, 2003)

• Trophoblastic markers proven to be not specific: HLA-G, NeuroD2, placenta GF (Tioa ML, 2007)

• 30 ml blood, CD105, CD141, CK: Hatt L et al, 2013; Shlutter et al 2014; Shlutter et al, 2015:



## ISET® TECHNOLOGY for CIRCULATING FETAL TROPHOBLASTIC CELLS



ISET by Rarecells: Patented combination of parameters allowing very sensitive and rapid isolation of CFC

## Workflow for ISET® *in vitro* assay of sensitivity and reproducibility with fixed cells counted by micropipetting

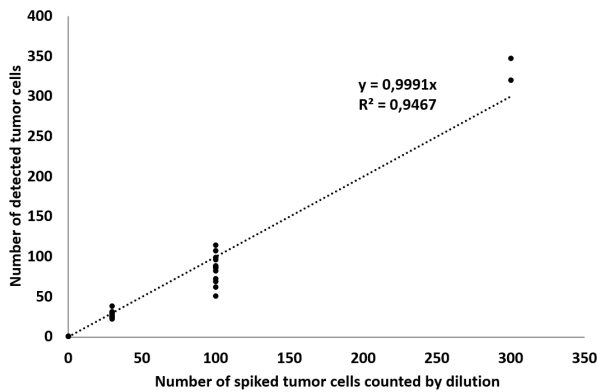
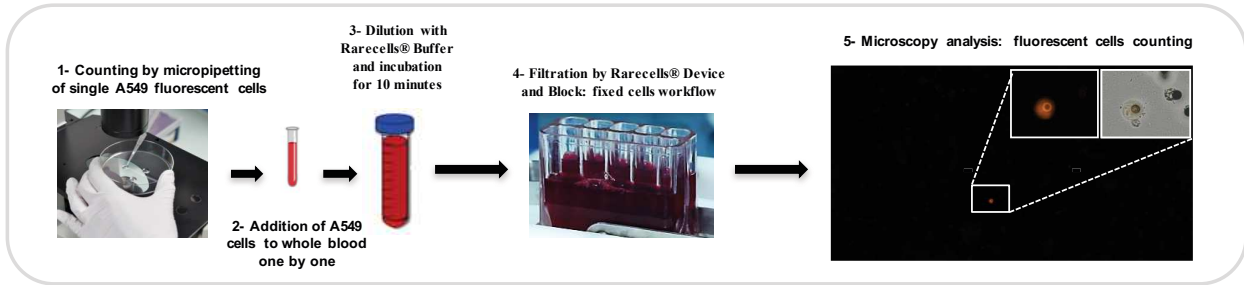


Table 2: Sensitivity and repeatability of the ISET® system (fixed cells)

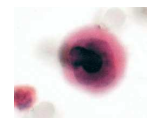
mL of blood processed	1 mL	5 mL	10 mL
Number of spiked tumor cells	2	2	2
	1	1	2
	2	2	2
Number of tumor cells detected by ISET®	2	2	2
	1	2	2
	2	2	2
	2	1	2
total found /spiked	10/12	10/12	12/12
Recovery success rate	83,3%	83,3%	100%

## Results

G. VONA, .....PATERLINI-BRÉCHOT. Enrichment and genetic analyses of fetal cells circulating in the maternal blood by the ISET technique and single cell microdissection : a non-invasive tool for early prenatal diagnosis. *Am J Pathol*, 160 : 51-58, 2002.



13 pregnant women  
11-12 WG  
Before CVS  
CFTC in all the mothers, Y and STR genotyping



C. BÉROUD, .....PATERLINI-BRÉCHOT. Prenatal diagnosis of Spinal Muscular Atrophy (SMA) by genetic analysis of circulating fetal cells. *The Lancet*, 361 :1013-4, 2003.



12 mothers at risk for baby with SMA  
10-12 WG before CVS  
Blind analysis vs CVS  
CFTC in all the mothers, correct diagnosis

A. SAKER,....., P. PATERLINI-BRÉCHOT. Genetic characterization of circulating fetal cells allows Non-Invasive prenatal diagnosis of cystic fibrosis. *Prenat Diagn*, 26 : 906-16, 2006.



12 mothers at risk for baby with CF  
10-12 WG  
Blind analysis vs CVS  
CFTC in all the mothers, correct diagnosis

H. MOUAWIA, ..... P. PATERLINI-BRECHOT. Trophoblasts enriched from maternal blood provide definitive genetic diagnosis in 63 consecutive fetuses at risk for Cystic Fibrosis or Spinal Muscular Atrophy, *Reproductive Biomedicine Online*, 2012



63 mothers at risk for baby with SMA or CF  
10-12 WG, before CVS  
Blind analysis vs CVS  
CFTC in all the mothers, correct diagnosis

PFEIFER I, .....P. PATERLINI-BRECHOT. Cervical Trophoblasts for non-invasive single cell genotyping and prenatal diagnosis, *Placenta*, 2015



21 pregnant women  
6 before CVS, 3 CF, 3 SMA, correct diagnosis  
15 before TOP  
PAP like sampling 8-12 WG  
2 to 12 trophoblasts in 2 out of 10 ml  
TC in all the mothers, correct diagnosis

# ISET Clinical validation study NI-PND of Cystic Fibrosis and Spinal Muscular Atrophy NECKER HOSPITAL BLIND PROTOCOL: Pr Benachi (obstetrician); Dr Bonnefont (geneticist); JP Jais (statistician)

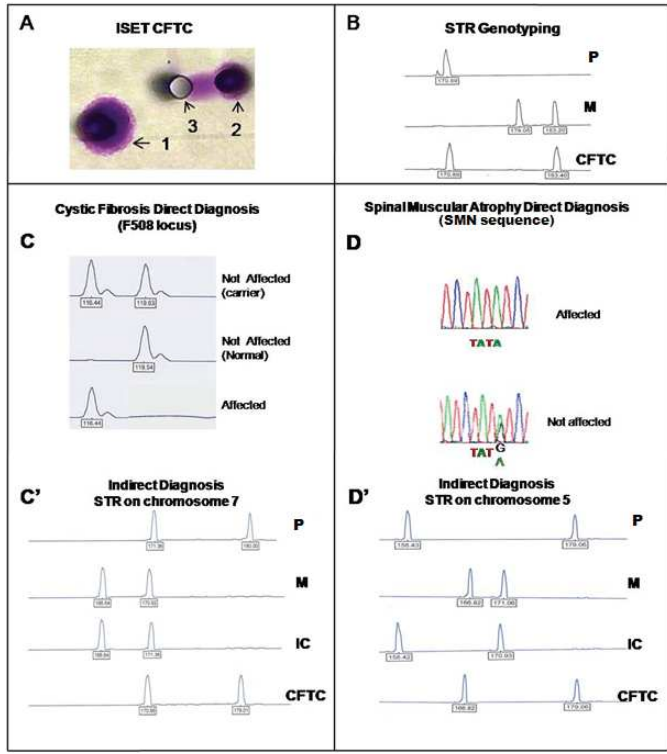
Table 2 Non invasive prenatal diagnosis for spinal muscular atrophy.

Couple no. (weeks of gestation)	STR markers	Microdissected cells (n)	CFTC (n)	CFTC result (n)		Test results	
				With gene deletion	Without gene deletion	NI-PND	Invasive PND
1 (11)	D16S339, D16S3018, D21S1435	13	6	0	5	NA	NA
2 (11)	D7S486, D16S3018, D21S1435	11	5	0	5	A	A
3 (10)	D7S486, D7S486, D7S323	11	5	0	5	NA	NA
4 (10)	D7S486, D21S1435, D21S1437	15	7	0	5	NA	NA
5 (9)	D16S339, C272, D21S1435, D16S2800	17	8	0	5	A	A
6 (9)	D16S339, D5S816, D21S1435	15	8	0	5	NA	NA
7 (8)	D21S1437, D17S200, D20S116	12	6	0	5	NA	NA
8 (11)	D16S339, D5S816, D21S1435	14	6	0	5	NA	NA
9 (11)	D16S339, D21S1435, D21S1437	12	7	0	5	NA	NA
10 (10)	D7S323, D16S339, D5S816	11	5	0	5	NA	NA
11 (9)	D7S486, D16S339, D21S1435	19	9	0	5	A	A
12 (11)	D7S486, D16S3018, D21S1437	16	7	0	5	NA	NA
13 (10)	D5S317, D16S339, D20S116	14	6	5	0	A	A
14 (11)*	D16S339, D21S1437, D20S116	11	5	0	5	NA	NA
15 (11)	D16S339, D21S1437, D20S116	12	6	0	5	NA	NA
16 (9)*	D16S339, D5S816, D20S116	12	7	0	5	NA	NA
17 (11)*	D16S339, D5S816, D21S1435	24	11	0	10	NA	NA
18 (10)*	D7S323, D16S339, D5S816, D17S200	21	10	0	10	NA	NA
19 (10)*	D16S339, D5S816, D17S200	25	12	0	10	NA	NA
20 (11)*	D16S339, D21S1437, D21S1435	21	5	10	0	A	A
21 (11)	D16S339, D21S1435, D21S1437	27	12	0	10	NA	NA
22 (10)	D16S339, D21S1437, D20S116	22	10	0	10	NA	NA
23 (10)*	D16S339, D5S816, D21S1435, D17S200	27	13	0	10	A	A
24 (10)	D16S339, D5S816, D21S1435	27	12	10	0	A	A
25 (11)*	D16S339, D5S816, D20S116	27	12	0	10	NA	NA
26 (9)*	D21S1437, D17S200, C272	23	11	0	10	NA	NA
27 (11)	D16S339, D21S1437, D21S1435	22	12	0	10	NA	NA
28 (10)	D16S339, D21S1435, D151171, D7S486	21	10	0	10	NA	NA
29 (11)	D21S1435, D7S486, D17S200	24	10	0	10	NA	NA
30 (11)	D16S339, D21S1435, D16S3018	22	10	0	10	NA	NA
31 (11)	D21S1435, D151171, D7S486	21	10	0	10	NA	NA
Total SMA		586	276	50	185		

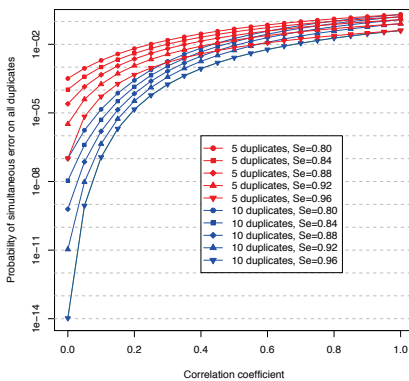
A - affected; C - carrier; CFTC - circulating fetal trophoblastic cells; CVS - chorionic-villus sampling; N - normal; NA - not affected; NI-PND - non-invasive prenatal diagnosis; STR - short tandem repeat.  
\*Cases redundantly tested by indirect diagnosis.

Couple no. (weeks of gestation)	STR markers	Microdissected cells (n)	CFTC (n)	CFTC result for F508del (n)			Test results	
				Homozygous	Heterozygous	Without F508del	NI-PND	Invasive PND
1 (11)*	D7S486, D16S339, D21S1435	14	7	0	5	0	C	C
2 (9)*	D7S486, D16S339, D21S1437	12	6	0	5	0	C	C
3 (11)	D16S339, D21S1435, D21S1437	16	7	0	0	5	H	N
4 (10)	D16S339, D21S1437, D21S1435	15	8	0	0	5	H	N
5 (10)	D16S339, D16S3018, D21S1435	14	6	0	0	5	N	N
6 (9)*	D7S486, D16S339, D16S2818	12	6	0	0	5	N	N
7 (11)	D16S339, D21S1437, D21S1435	19	9	0	5	0	C	C
8 (11)	D16S339, D21S1437, D21S1435	12	7	0	0	5	C	C
9 (10)*	D7S486, D16S3018, D21S1435	12	6	5	0	0	A	A
10 (9)*	D7S486, D7S486, D7S323	11	5	10	0	0	A	A
11 (10)*	D7S486, D21S1435, D21S1437	13	6	0	5	0	C	C
12 (10)	D7S486, D7S486, D7S323	17	8	0	0	5	H	N
13 (11)*	D7S486, D21S1435, D16S3018	13	7	10	0	0	A	A
14 (11)*	D7S486, D21S1435, D21S1435	13	6	5	0	0	A	A
15 (11)	D7S486, D16S339, D21S1435	11	5	0	0	5	C	A
16 (10)*	D7S486, D16S339, D16S2818	15	7	5	0	0	A	A
17 (10)	D16S339, D21S1437, D7S486	23	11	0	0	10	N	N
18 (11)	D16S339, D21S1435, D151171	27	12	0	10	0	C	C
19 (10)	D16S339, D7S486, D5S816	24	11	0	0	10	C	C
20 (9)*	D16S339, D5S816, D7S486	26	12	0	0	10	N	N
21 (11)	D16S339, D16S3018, D21S1435	21	10	0	10	0	C	C
22 (11)	D16S339, D21S1437, D5S816	24	11	0	10	0	C	C
23 (11)	D16S339, D21S1435, C272	21	10	0	10	0	C	C
24 (11)*	D7S486, D16S3018, D16S2818	22	10	0	10	0	H	N
25 (11)	D16S339, D5S816, D21S1437	21	11	0	0	10	N	N
26 (10)*	D7S486, D16S339, D20S116	30	13	10	0	0	A	A
27 (9)*	D7S486, D7S486, D21S1437	23	11	0	0	10	N	N
28 (10)	D7S323, D21S1435, D16S339	25	12	10	0	0	A	A
29 (11)*	D7S486, D16S339, D5S816	25	12	10	0	0	A	A
30 (11)	D21S1435, D7S486, D5S816	21	10	0	0	10	C	C
31 (11)*	D7S323, D21S1435, D7S486	26	12	10	0	0	A	A
32 (10)	D7S323, D21S1435, D7S486	27	12	10	0	0	A	A
Total CF		606	285	55	105	80		

A - affected; C - carrier; CFTC - circulating fetal trophoblastic cells; CVS - chorionic-villus sampling; N - normal; NI-PND - non-invasive prenatal diagnosis; STR - short tandem repeat.  
\*Cases redundantly tested by indirect diagnosis.  
\*One parent with non-F508del mutation.  
\*Both parents with non-F508del mutation.



## Successful clinical validation - Mouawia et al, RBMO 2012



Effect of circulating fetal trophoblastic cell replicated tests on the overall reliability of the diagnostic procedure.

When using 10 CFTC replicates, the protocol of independent diagnostic analysis (correlation = 0), virtually never gives rise to a global (clinical) mistake at an error rate of 10%, i.e. the probability of global diagnostic error rate is 1 in 10 billion. With five CFTC replicates, this extraordinary performance can be reached if the error rate is 1% or less.

**SMA + CF: total (31+32) = 63 pregnant women**

**Consecutive cases**

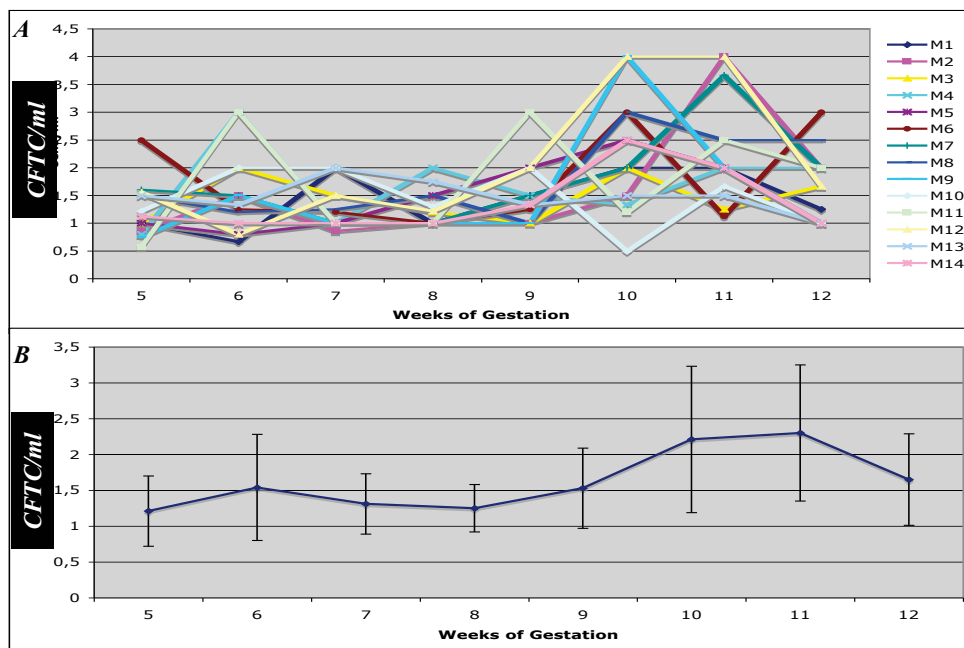
**1191 microdissected cells: 475 CFTC**

**7 + 7 affected fetuses  
as compared to CVS  
Sensitivity: 100%  
Specificity: 100%**

# Kinetics of Circulating Fetal Trophoblastic Cells in maternal blood after In Vitro Fertilization (4th to 12th WG) (collab. Prof R Frydman):

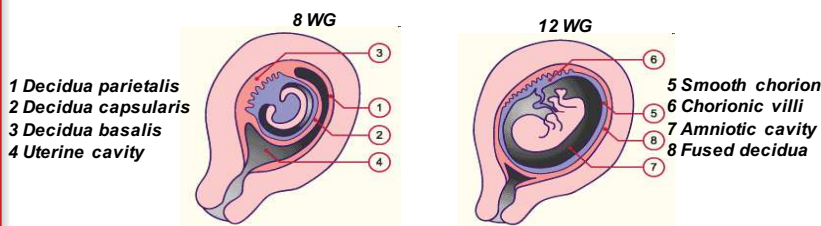
## CFTC start to circulate at the 5th WG

Total  
473 CFTC



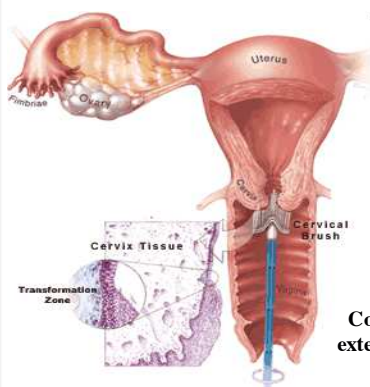
# Isolation of trophoblasts from the cervix

Transcervical cells (TCC): inner part of the cervix and lower pole of the uterine cavity (Schettles 1971)



## TCC sampling methods:

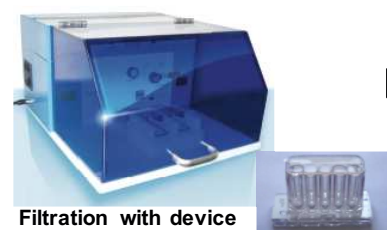
- Intrauterine lavage
- Endocervical lavage
- Endocervical mucus aspiration
- Endocervical sampling by a cytobrush



Collection of cells from external part of the cervix



1:10 dilution



trophoblastic cells on filter

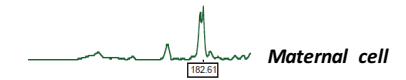


## Non-invasive isolation of trophoblasts from the cervix 2ml out of 10 ml analyzed - Pfeifer I 2015

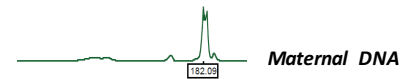
Couple	Term of pregnancy (WG)	Informative STR marker	Cytotrophoblasts/Syncytiotrophoblasts* - NIPD**	N° of microdissected cells
1(CF)	12	D7S486/D7S523	4 - carrier	10
2(CF)	12	D7S523	6 - carrier	12
3(CF)	12	D16S539/D7S523	10 - carrier	19
4 (SMA)	12	D5S816/D21S1437	6 – not affected	13
5 (SMA)	12	D21S1435	10 – not affected	21
6 (SMA)	12	D16S539/D7S523	6 – not affected	13
7#	12	D5S816/D21S1437	5	11
8#	12	D16S539	3/2	9
9#	11	D16S539/D5S816	4/2	10
10#	12	D21S1435	10	21
11#	12	D21S1435	6	14
12#	12	D16S3018	7	13
13#	12	D21S1435/D7S523	6	14
14#	12	D16S539/D5S816	2	6
15#	12	D21S11	8/2	16
16#	12	D16S539/D21S1435	12	21
17#	9	D16S3018/D5S615	6	12
18#	9	D5S615/D16S539	4	9
19#	8	D16S539/D5S816	4	10
20#	11	D16S539/D21S11	3	7
21#	8	D5S615/D5S816	3/1	8



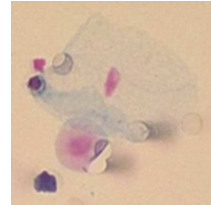
Maternal cell



Maternal cell



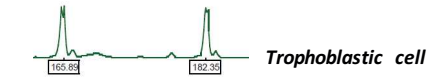
Maternal DNA



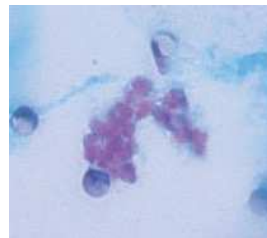
Trophoblastic cell



Paternal DNA



Trophoblastic cell



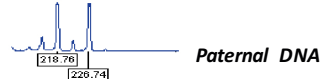
Syncytiotrophoblast



Syncytiotrophoblast

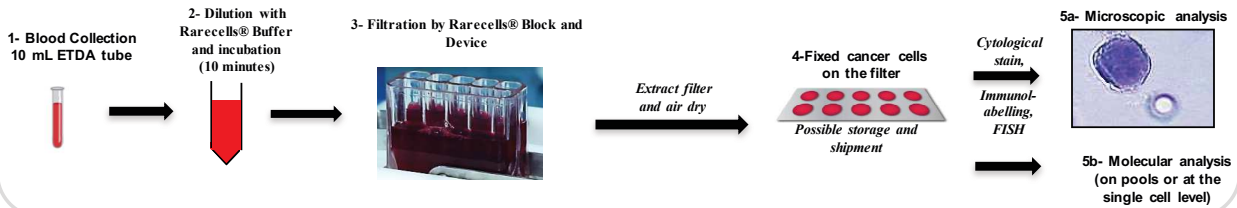


Maternal DNA

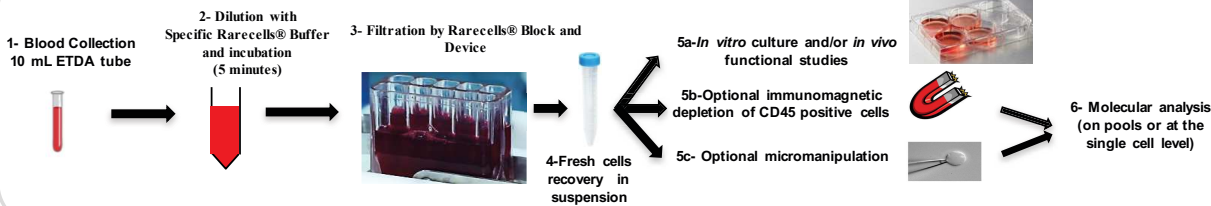


Paternal DNA

### A- ISET® filtration workflow for enrichment and analysis of fixed circulating rare cells



### B- ISET® filtration workflow for enrichment and analysis of fixed/live circulating rare cells



- Workflow for ISET® *in vitro* assay of sensitivity and reproducibility with fixed/live cells counted by micropipetting

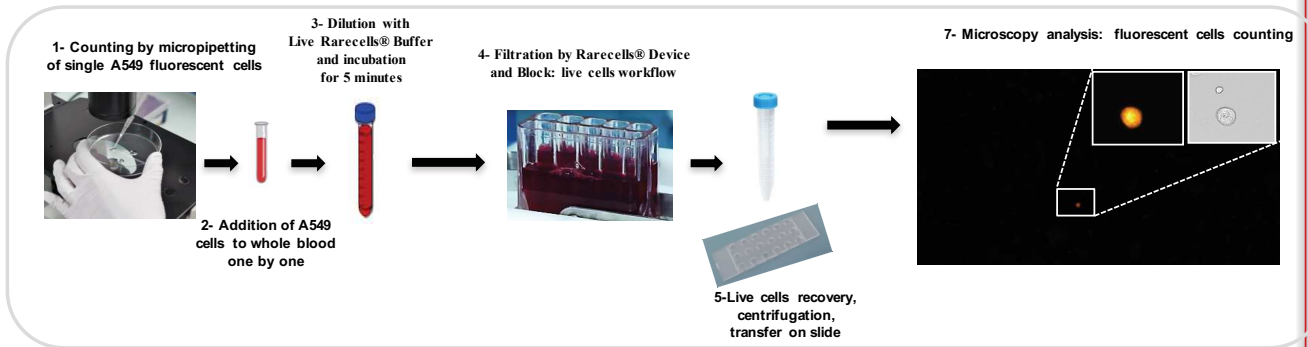


Table 5. Sensitivity and repeatability of the ISET® system for live tumor cells

Cell line	A549				LNCaP	
Number of spiked cells	1	3	5	1	1	5
Number of tumor cells detected by ISET®	1	3	4	9	1	4
	1	3	5	6	1	4
	1	2	4	8	0	5
	1	3	4	9	1	3
	1	2	3	10	1	5
	1	3	4	8		
Total detected cells / spiked cells	6/6	16/18	24/30	50/60	4/5	21/25
Recovery rate (SEM)	100% (0)	88.8% (7)	80% (5.2)	83.3% (5.6)	80% (20)	84% (7.5)

SEM, Standard error of mean

Sensitivity of the ISET® system for dual collection of live rare cells followed by CD45-immunomagnetic depletion

Blood volume (mL)	1
Number of spiked cells	10
Number of rare cells detected by ISET®	4
	3
	3
Total detected cells / spiked cells	21/60
Recovery rate (SEM)	35% (5.2)

SEM, standard error of mean; two independent experiments in triplicates

**> 30  
CFTC per  
10 ml blood**

**30% = > 10  
CFTC  
WGA  
genotyping  
(single cells or pooling)  
NGS**

## Acknowledgements

**INSERM  
and Laboratoire de Biochimie A,  
Hôpital Necker-Enfants Malades, Paris,  
France, Université Paris Descartes,**

**Hussein Mouawia  
Ali Saker  
Ingrid Pfifer  
Lucile Broncy  
Patrizia Paterlini Bréchet**

**Department of Pharmaceutics  
University of Ghent  
Belgium**

**Philip Van Nieuwerburgh  
Lieselot Deleye**

**FASTERIS**  
MEMBER OF THE INNOVATION NETWORK  
**Geneva - Switzerland  
Dr Laurent Farinelli  
Dr Magne Osteras**

**Service de Gynécologie-Obstétrique,  
Hôpital Antoine Béchère, Clamart, France;  
Alexandra Benachi**

**Laboratoire de Génétique Médicale, Hôpital  
Necker-Enfants Malades, Paris, France;**

**Jean-Paul Bonnefont**

**Service de Gynécologie-Obstétrique,  
Hôpital Foch, France;**

**René Frydman**

**Biostatistique, Université Paris Descartes,  
Jean-Philippe Jais,  
Unité de Recherche Clinique Paris Ouest, Hôpital  
Ambroise-Paré AP-HP, Boulogne Billancourt  
Laurence Bussièrès**